




# The Open Dentistry Journal

Content list available at: <https://opendentistryjournal.com>



## RESEARCH ARTICLE

### Antibacterial Behavior of Silver Diamine Fluoride, Sodium Hypochlorite and Ozone Gel on *Enterococcus faecalis* in Root Canal of Deciduous Teeth

Naimeh Teimoori<sup>1</sup> , Fatemeh Yegane-Sefidan<sup>2</sup>, Roghayeh Nouri<sup>2</sup>, Leila Erfanparast<sup>1,\*</sup>, Yashar Rezaei<sup>3</sup> and Masoomeh Vatandoost<sup>1</sup>

<sup>1</sup>Department of Pediatric Dentistry, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran

<sup>2</sup>Faculty of Medicine, Department of Microbiology, Tabriz University of Medical Sciences, Tabriz, Iran

<sup>3</sup>Department of Dental Biomaterial, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran

#### Abstract:

#### Introduction:

*Enterococcus faecalis* (*E. faecalis*) has been reported as one of the most important causes of failed endodontic treatments. Various antibacterial agents have been suggested to remove *E. faecalis* from the root canal. We aimed to investigate and compare the antibacterial efficacy of silver diamine fluoride (SDF), Sodium hypochlorite (NaOCl) and ozone gel on *E. faecalis* in the root canal.

#### Materials and Methods:

This study involved 60 extracted roots of molar teeth with a single root canal to generate a 6-week-old biofilm for evaluating antibacterial effects. After teeth decorating and removing the smear layer using sodium hypochlorite (2.5%) and EDTA (17%), roots were sterilized by autoclave. Under sterile conditions, 1 ml of the suspension containing *E. faecalis* bacteria was transferred to each of the microtubes containing teeth. The samples were divided into four groups: exposed to NaOCl (2.5%) as a positive control, exposed to liquid ozone (25 ppm), exposed to SDF (3.8%), and exposed to normal saline as a negative control group. Then, the colony forming unit (CFU) was counted in the studied groups. Kruskal-Wallis test and Dunn-Bonferroni post-hoc analysis were carried out for comparison of CFU in the studied groups.

#### Results:

The mean of CFU were 17846, 920, 234, and 336 for saline, ozone, NaOCl, and SDF, respectively. There were significant differences in CFU in the comparison of (NaOCl-Ozone), (NaOCl-saline), (SDF-saline), and (ozone-saline) ( $P < 0.05$ ). However, there was no significant difference for CFU in the comparison of NaOCl-SDF ( $P = 0.570$ ).

#### Conclusion:

The study findings showed that NaOCl (2.5%) and SDF (3.8%) were more effective against the 6-week-old *E. faecalis* biofilms than ozone and saline.

**Keywords :** Antibacterial agents, *Enterococcus faecalis*, Root Canal Therapy, Deciduous Teeth, Silver Diamine Fluoride, Endodontic Treatment.

#### Article History

Received: May 10, 2023

Revised: August 05, 2023

Accepted: August 23, 2023

## 1. INTRODUCTION

Appropriate root treatment maintains the function of deciduous teeth with severe decay [1]. The achievement of a good endodontic treatment in primary teeth is mainly governed by the diminution or removal of bacteria from root canals by antibacterial agents [2]. Although many materials have been suggested for washing and disinfecting the root canal of perma-

nent teeth, the growing, anatomical and physiological variances between deciduous and permanent teeth cause differences in the criteria of the materials used in the stages of root canal pulpectomy [3].

Studies on teeth with failed endodontic treatment have often reported the prevalence of *Enterococcus faecalis* (*E. faecalis*), a facultative gram-positive bacterium, in persistent or secondary infections [4]. The invasion of dentin tubules, adaptation to the unfavorable conditions created inside the root canal, and resistance to irrigations and drugs inside the canal

\* Address correspondence to this author at the Department of Pediatric Dentistry, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran; Tel: 041-33355965; Fax: 041- 33346977; E-mail: leilaerfan@gmail.com,

are among the reasons for the resistance of this bacteria. Biofilm maturation occurs over time through the mineralization of its structure, and *E. faecalis* biofilm maturation occurs within six weeks. Then, six weeks is considered as the time index of mature *E. faecalis* biofilm [5].

The use of sodium hypochlorite (NaOCl), chlorhexidine (in gel or liquid forms), silver diamine fluoride (SDF), and ozone are among the most effective ways to remove *E. faecalis* from the root canal [6]. NaOCl is the most common agent used for root canal disinfection in concentrations of 0.5 to 5.25% [7]. In most studies, there is no significant difference in the antimicrobial effect of 5% and 2.5% NaOCl [7, 8]. Sodium hypochlorite has disadvantages such as an unpleasant taste, high toxicity, burning surrounding tissues, and inability to remove the smear layer. Another disadvantage of using NaOCl is its unpleasant smell [6, 9].

Due to the disadvantages of NaOCl, the use of other antibacterial materials such as chlorhexidine, ozone, and SDF was also suggested in the previous studies [10 - 12]. Ozone, as a natural gas, is a strong selective oxidizer. The main foundation of ozone therapy is the rapid decomposition of ozone in water and releases a form of oxygen that may oxidize cells, thus, it has the potential to have an antimicrobial effect without causing drug resistance [13, 14]. Currently, ozone gas is used for adult root canal disinfection and has been demonstrated to be less cytotoxic than 2.5% NaOCl [2].

SDF, as a strong anti-cariogenic material, shows high fluoride release and the ability to remineralize the tooth surface and strengthens its resistency [15]. Although the silver in SDF has an antimicrobial role, its antibacterial effect against root pathogens is poorly understood [16]. Unlike chlorhexidine, SDF has not been shown to be cytotoxic, so it is considered a potential root irrigation [11]. There is limited evidence for the assessment of the antimicrobial action of NaOCl, SDF, and ozone against root pathogens. Therefore, the current study aimed to investigate the antibacterial efficacy of SDF, NaOCl and ozone gel on *E. faecalis* in the root canal of deciduous teeth.

## 2. MATERIALS AND METHODS

### 2.1. Tooth Samples

Sixty-three extracted mesial roots of mandibular molar teeth with a single root canal were used to generate a 6-week-old biofilm model for evaluating the antibacterial efficacy of silver diamine fluoride (SDF), sodium hypochlorite (NaOCl) and ozone gel against *E. faecalis* in comparison with normal saline. The sample size was determined based on the earlier study [17] by considering the mean $\pm$ SD ( $1.29 \times 10^8 \pm 0.36$ ) of colony numbers,  $\alpha = 0.05$ , and power = 0.80.

### 2.2. Eligibility Criteria

#### 2.2.1. Inclusion Criteria

The extracted mesial roots of mandibular deciduous molar teeth due to orthodontics or over-retained.

#### 2.2.2. Exclusion Criteria

The teeth with previous pulp treatment, previous restorative treatments, teeth with any previous dental treatment, and teeth with external and internal root resorption. The teeth with apical resorption of more than 1/3 of root length were also excluded from the study.

### 2.3. Preparation of Dentin Specimens

#### 2.3.1. Preparation of Roots

After debridement of teeth, the samples were sterilized in an autoclave at 121 °C for 20 min. The roots were sectioned off, applying a diamond disc (D&Z, Diamant, Germany). Then, to split the roots using a low-speed saw (IsoMet 2000 Precision Saw; IsoMet, Buehler, IL) rotating at 1,000 rpm in a buccolingual direction under water cooling was done. Then, mesial roots were selected. Roots with a length of 7 mm were prepared.

#### 2.3.2. Preparation of Canals

The pulp tissue of the canals was removed using a barbed broach (Dentsply, Maillefer, Ballaigues, Switzerland) and the instrumentation of the canals was done by the step-back method.

To remove the smear layer, one ml of 2.5% sodium hypochlorite (Hypo-Endox, Morvabon, Iran) for 1 minute and then one ml of 17% EDTA (Pulpdent Corp, MA, US) for 1 minute were used. Lastly the washing step of roots was done for 1 minute with sterile saline. The samples were then sterilized by an autoclave at a temperature of 121 °C for 15 minutes with a pressure of 15 pounds per square inch. Lastly the washing step of roots was done for 1 minute with sterile saline [18].

#### 2.3.3. Bacterial Inoculation

The standard strain of *Enterococcus faecalis* (ATCC 29212) was cultured under sterile conditions on Brain Heart Infusion Agar (QUELAB, Canada) and incubated for 24 hours at 37 °C. After the incubation period, the half McFarland ( $1.5 \times 10^8$  CFU/ML) suspension of the strain was prepared in Brain heart infusion Broth (BHI Broth) (QUELAB, Canada). Under sterile conditions, one milliliter of the prepared concentration was transferred to each of the microtubes containing teeth and incubated for 24 hours at 37 °C. In order to form a mature biofilm for 6 weeks, the suspension inside the micro-tubes was replaced with a new microbial suspension every other day for the purpose of simulating the real conditions of the teeth in the mouth. After 6 weeks, 3 samples were randomly examined to confirm the formation of biofilm under an electronic microscope (LEO s-440i, UK).

#### 2.3.4. The Study Groups

A total of 60 samples were randomly assigned into 4 groups (A, B, C, and D), including 15 samples in each group. The samples were exposed to 0.5 ml of 2.5% sodium hypochlorite as a positive control group (group A), liquid ozone with a concentration of 25 ppm (group B), silver diamine fluoride (SDF) 3.8% (group C), and normal saline as a negative control group (group D), for 1 min. Finally, the canals

of all samples in all 4 groups were washed with 10 ml of sterile serum and dried using an aseptic paper cone (DATA, China).

### 2.3.5. The Number of Colonies: the Colony Forming Unit (CFU)

Sampling was done by Gates Glidden number 4. Then, each of the dental debris was dissolved in 500 microliters ( $\mu\text{L}$ ) of sterile normal saline and the dilution method was used to count the colony forming unit (CFU) on the same day. The microtube contained 5  $\mu\text{L}$  dissolved debris and 45  $\mu\text{L}$  of normal saline with three concentrations of  $10^{-1}$ ,  $10^{-2}$ , and  $10^{-3}$  were prepared. Then, 10  $\mu\text{L}$  of each prepared concentration was transferred separately to solid agar culture media and spread on the surface of the plate.

After counting the CFU, the obtained number was multiplied by the reciprocal of the selected concentration. Considering that the volume removed for cultivation on the surface of the plate to count the CFU was 10  $\mu\text{L}$ , the obtained number was again multiplied by 100 to obtain the number of CFU in one milliliter (1000  $\mu\text{L}$ ).

### 2.4. Statistical Analysis

For data analysis, we used SPSS software (version 19.0, Chicago, IL, USA). The Kolmogorov-Smirnov test was passed out for testing the normality of data. Descriptive statistics, including mean, Standard Deviation (SD), median, and percentile, were used to present the number of colonies in the study groups. Kruskal-Wallis test was used to compare the between-groups comparison of colonies number since the number of colonies had non-parametric distribution. Dunn-Bonferroni was carried out for post-hoc analysis. The significant level was in P-value  $<0.05$ .

### 3. RESULTS

Overall, 60 extracted mesial roots of mandibular molar teeth were collected; then samples were irrigated with 2.5% sodium hypochlorite (group A), liquid ozone 25 ppm (group B), 3.8% SDF (group C), and normal saline (group D); The step followed by culturing the bacteria that survived after using irrigations, the antimicrobial properties were determined by counting the number of colonies formed and then calculating the number of colonies in one milliliter.

Table 2 shows the mean, median, and descriptive statistics of colonies in the study groups. The mean of CFU in the saline group had a maximum amount, while it had a minimum amount in the NaOCl group. The mean CFU were 17,846, 920, 234, and 336 in saline, ozone, NaOCl, and SDF, respectively. However, the median CFU was zero for NaOCl, and SDF. The mean and SD, and percentiles of the number of colonies are presented in Table 1.

Table 2 and Fig. (1) indicate the comparison of the number of colonies in the studied groups. There was a significant difference in the number of colonies between groups ( $P=0.001$ ).

Table 3 shows Dunn-Bonferroni post-hoc analysis of the CFU (pairwise comparison of substances with control groups (NaOCl and saline)). There was a significant difference in the

number of colonies (CFU) in two-by-two comparison of groups NaOCl-ozone, NaOCl-saline, SDF-saline, and ozone-Normal saline (all  $P < 0.05$ ). However, there were no significant differences for the number colonies in comparison to NaOCl-SDF groups ( $P=0.570$ ).

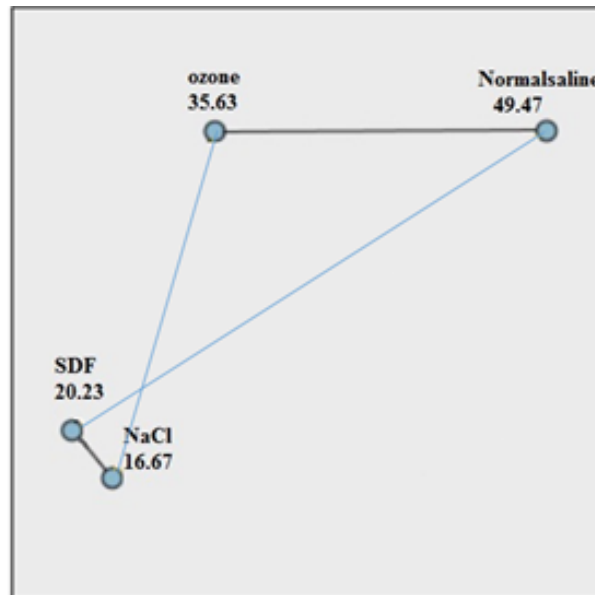


Fig. (1). Between-group comparison of the number of colonies.

### 4. DISCUSSION

In the current *in-vitro* study, the antimicrobial effect of 3.8% SDF, 25 ppm ozone gel, 2.5% sodium hypochlorite and normal saline was investigated on the six-week biofilm of *E. faecalis* bacteria. The study findings demonstrated that 2.5% NaOCl and 3.8% SDF for 1 minute were the most effective disinfectant protocols against the six-week biofilm of *E. faecalis* bacteria. Our analysis found that there is no significant difference regarding disinfectant characteristics 2.5% NaOCl in comparison with 3.8% SDF. Furthermore, in several samples, they were able to completely remove 100% of the bacterial biofilm. On the other hand, although the antibacterial effect of 25 ppm ozone was more effective than normal saline, however, normal saline and ozone showed the least effect on the six-week biofilm of *E. faecalis* bacteria.

*E. faecalis* is one of the most common microorganisms in infected root canals of deciduous teeth, and is associated with pulp necrosis [19]. Since the bacteria grown in the biofilm are more resistant to antimicrobial treatment than the planktonic bacteria, and the full maturation of the *E. faecalis* biofilm by its mineralization occurs within six weeks [20, 21]. It is difficult to remove the adult mineralized biofilm of *E. faecalis* by conventional methods and ultimately leads to resistant and persistent root canal infections [22]. Therefore, the antibacterial protocols were evaluated in this study on the six-week biofilm of *E. faecalis*. In order to form a mature biofilm for 6 weeks, we replaced the suspension inside the microtubes with a new microbial suspension every other day.

Overall, root canal treatment is considered the most effective way to eliminate inflammation and infection in the

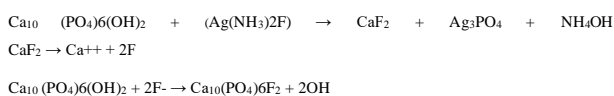
root canal, and sometimes, the presence of chronic infection in the root canal may lead to treatment failure [23]. It is almost impossible to remove bacteria using physical methods due to the complications and limitations of the root canal system. The minimum goal of root canal treatment is to reduce the number of bacteria to a level lower than the necessary value for the reproduction of the disease [24]. Irrigations have a major role in pediatric endodontics due to their complex structure and features, such as internal connections and horizontal anastomoses seen in primary teeth [25].

In group A, the samples were exposed to 0.5 ml of 2.5% sodium hypochlorite for 1 minute. Currently, sodium hypochlorite is the most common substance that is used to disinfect the root canal, and it is used in recommended concentrations of 0.5 to 5.25% [7]. There is no agreement regarding the ideal concentration required with sodium hypochlorite to remove bacteria from the root canal. Likewise, the study of Reyhani *et al.* showed that 2.5% and 5% sodium hypochlorite completely remove the six-week-old biofilm of *E. faecalis* bacteria. However, as the age of the biofilm increases, its resistance increases compared to 1% sodium hypochlorite [8].

In group B, the samples were exposed to 0.5 ml of liquid ozone with a concentration of 25 ppm for 1 minute. Ozone gel is in oil form in the refrigerator and liquid form at room temperature. Ozonized olive oil gel was chosen for the study because, unlike ozone gas, it does not require expensive devices and the concentration of molecules of ozone is higher and the stability of its composition is also longer [26]. Ozone rapidly breaks down in the water and releases a form of oxygen that may oxidize cells, thus potentially having an antimicrobial effect without causing drug resistance [27]. Regarding the antibacterial effect of ozone, it is able to dissolve debris inside the canal, disinfect parts of the root canal that are not accessible by mechanical tools, be effective against root canal pathogenic microorganisms such as *E. faecalis*, *Candida albicans*, *Peptostreptococcus*, and penetrate through the apical foramen to bone, repair, reconstruction, and wound healing and surrounding bone [15, 28].

In group C, the samples were exposed to 0.5 ml of 3.8% silver diamine fluoride for 1 minute. The SDF was used as an anti-caries agent (38% SDF), which is especially effective in children's dentistry. The SDF is efficient and safe, and it is biocompatible, especially in lower concentrations [29 - 31]. 3.8% silver diamine fluoride solution was developed for use as an irrigation solution in root canal treatment [32].

One of the mechanisms of the anti-caries function may be caused by the products produced from the reaction between SDF and mineral parts of the tooth. The basic chemical reaction is shown below:



The existing fluoride strengthens the resistance of the dentin against the acid, which leads to a decrease in the penetration of the acid into the inner dentin [33]. Also SDF may lead to the hardening of the affected dentin by reacting

with dental hydroxyapatite (HA) and releasing calcium fluoride and silver phosphate [34].

The silver of SDF has an antimicrobial role. The antimicrobial role of silver diamine fluoride is related to the formation of organometallic complexes inside bacteria cells. These substances cause the deactivation of enzymes by blocking the electron transport system in bacteria. Also, they cause mutation and death of these cells by interfering with the DNA of bacteria [16].

However, in group D, we used normal saline as a negative control group for comparison of the antibacterial effect in the studied groups.

Generally, the results of our study regarding the antibacterial effect of 3.8% SDF and 2.5% sodium hypochlorite are in line with the results of previous studies.

Al-Madi *et al.* reported that NaOCl indicated the highest percentage of dead cells, with 62.26%. The SDF group demonstrated a significantly higher percentage of dead cells (57.39%) than the 2% CHX and normal saline groups. They found that SDF had a higher antimicrobial effect than 2% CHX against *E. faecalis* biofilms [35]. Likewise, Hiraish *et al.* found that both NaOCl and SDF were effective against *E. faecalis* biofilms, with no significant difference in reduction of microorganisms for the same exposure time [32]. According to Chaudhari *et al.* NaOCl had a high effect as a root canal irrigant, followed by SDF and bioactive glass nanoparticle. In that study, the chitosan nanoparticles showed the least antimicrobial effect [36].

Some previous studies evaluated the antibacterial effect of ozone more than what was shown in the current study [13, 37].

In a study conducted by Boch *et al.* the highest bacterial reduction was found in NaOCl group (99.98%), while EDTA decreased bacteria by 80.64%. Combination of NaOCl and ozone eliminated 99.95% of the bacteria. Combination of EDTA and ozone eradicated *E. faecalis* up to 91.33% [13]. As a result, ozone gas can dramatically reduce the biofilm of *E. faecalis* bacteria. In our study, liquid ozone did not have a successful effect in reducing *E. faecalis*. Possible reasons include the following: 1- Gaseous ozone was used in the above study, while liquid ozone was used in our study. 2- The higher concentration of ozone gas used in Boch *et al.* study compared to the current study 3- Younger biofilm of *E. faecalis* bacteria used in their studies compared to the current study.

The findings of the current study are in line with a systematic review by Saliva *et al.* demonstrated that ozone therapy has meaningfully less microbial effect than NaOCl. They showed that ozone is ineffective in increasing the antimicrobial effect of NaOCl. This finding was powerfully related to the protocol used: the dose, the time and the bacterial strain [38]. However, according to the previous findings, normal saline had the least antimicrobial effect in the present study [39, 40].

## 5. LIMITATIONS OF THE STUDY

The current study was an *in vitro* study and its results need to be approved by *in vivo* or clinical studies. Some issues in sampling could have affected the outcomes. The pure *E. faecalis* suspension was used in this study, but studies should

**Table 1. Descriptive statistics of the number of colonies by the study groups.**

Variables	Study Groups (n=60)			
	Normal Saline (n= 15)	Ozone (n= 15)	Hypochlorite Sodium (NaCl) (n= 15)	SDF* (n= 15)
Mean	17,846	920	234	336
Standard Deviation (SD)	34004	837	576	637
Median	3400	600	0.0	0.0
Percentiles	%25	1200	0.0	0.0
	%50	3400	600	0.0
	%75	20,000	1,400	120

Note: \* 3.8% Silver Diamine Fluoride

**Table 2. Between-group comparison of the number of colonies.**

Groups	Mean Rank	P- value*
Normal saline	49.47	0.001
Ozone	35.63	
Hypochloritesodium	16.67	
SDF	20.23	

Note: \*Kruskal-wallis

**Table 3. Post-hoc analysis of the number of colonies by the study groups.**

Pairwise Comparison of the Groups	Test Statistics	P-value*
NaCl-SDF	-3.56	0.570
NaCl-Ozone	18.96	0.03
NaCl-Normalsalin	32.80	0.001
SDF-Normalsalin	29.23	0.001
Ozone-Normalsalin	13.83	0.028

Note: \*Dunn-Bonferroni

be done concentrating on polymicrobial biofilms rather than individual microorganisms. Some studies reported that SDF may show staining issues after the use of 38% SDF. The concentration of SDF which is used in this study as a root canal irrigator (3.8%), represents a 1:10 dilution of the concentration used for arresting caries (38%). While this subject is not estimated to advance esthetic worries in the case of using this material in molar teeth, this issue should be evaluated in future studies.

## CONCLUSION

The study findings showed that 2.5% NaOCl and 3.8% SDF for 1 minute were more effective against the six-week biofilm *E. faecalis* biofilms than ozone and saline. We found no significant difference regarding disinfectant characteristics of NaOCl 2.5% in comparison with SDF 3.8% in this study.

## CLINICAL SIGNIFICANCE

Due to some disadvantages of NaOCl such as burning surrounding tissues and an unpleasant smell, the usage of SDF can be a better clinical option in root canal of deciduous teeth.

## LIST OF ABBREVIATIONS

SDF	=	Silver Diamine Fluoride
NaOCl	=	Sodium Hypochlorite
CFU	=	Colony Forming Unit
SD	=	Standard Deviation

## ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study was approved by ethics committee of Tabriz University of Medical Sciences to number: IR.TBZMED.REC.1401.419

## CONSENT FOR PUBLICATION

Not applicable.

## AVAILABILITY OF DATA AND MATERIALS

The data and supportive information are available within the article.

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

## FUNDING

This study has been funded by the Vice-Chancellor for Research (VCR) of Tabriz University of Medical Sciences (69561).

## ACKNOWLEDGEMENTS

Declared none.

## REFERENCES

- [1] Ghivari SB, Bhattacharya H, Bhat KG, Pujar MA. Antimicrobial activity of root canal irrigants against biofilm forming pathogens :An *in vitro* study. *J conserv dent* 2017; 20: 147. [PMID: 29279615]
- [2] Chandra SP, Chandrasekhar R, Uloopi KS, Vinay C, Kumar NM. Success of root fillings with zinc oxide-ozonated oil in primary molars: preliminary results. *Eur Arch Paediatr Dent* 2014; 15(3): 191-5. [http://dx.doi.org/10.1007/s40368-013-0094-8] [PMID: 24197970]
- [3] Trairatvorakul C, Chunlasikaiwan S. Success of pulpectomy with zinc oxide-eugenol vs calcium hydroxide/iodoform paste in primary molars: A clinical study. *Pediatr Dent* 2008; 30(4): 303-8. [PMID: 18767509]
- [4] Swapnil S, Sharma A, Shah N, Mandlik J, Ghogare A. Antimicrobial efficacy of triphala and curcumin extract in comparison with calcium hydroxide against *E. faecalis* as an intracanal medicament an study. *Paripex Indian J Res* 2017; 6: 876-9.
- [5] Kayaoglu G, Ørstavik D. Virulence factors of enterococcus faecalis: Relationship to endodontic disease. *Crit Rev Oral Biol Med* 2004; 15(5): 308-20. [http://dx.doi.org/10.1177/154411130401500506] [PMID: 15470268]
- [6] Joy Sinha D, D S Nandha K, Jaiswal N, Vasudeva A, Prabha Tyagi S, Pratap Singh U. Antibacterial effect of *Azadirachta indica* (neem) or *Curcuma longa* (turmeric) against *Enterococcus faecalis* compared with that of 5% sodium hypochlorite or 2% chlorhexidine *in vitro*. *Bull Tokyo Dent Coll* 2017; 58(2): 103-9. [http://dx.doi.org/10.2209/tdcpublication.2015-0029] [PMID: 28724858]
- [7] Dutner J, Mines P, Anderson A. Irrigation trends among american association of endodontists members: A web-based survey. *J Endod* 2012; 38(1): 37-40. [http://dx.doi.org/10.1016/j.joen.2011.08.013] [PMID: 22152617]
- [8] Frough-Reyhani M, Ghasemi N, Soroush-Barhaghi M, Amini M, Gholizadeh Y. Antimicrobial efficacy of different concentration of sodium hypochlorite on the biofilm of *Enterococcus faecalis* at different stages of development. *J Clin Exp Dent* 2016; 8(5): 84-e480. [http://dx.doi.org/10.4317/jced.53158] [PMID: 27957257]
- [9] Ismail S, Adyanthaya A, Sreelakshmi N. Intracanal irrigants in pediatric endodontics: A review. *Int J Appl Dent Sci* 2017; 3: 246-51.
- [10] Salomão PMA, Oliveira FA, Rodrigues PD, et al. The cytotoxic effect of TIF4 and NaF on fibroblasts is influenced by the experimental model, fluoride concentration and exposure time. *PLoS One* 2017; 12(6): e0179471. [http://dx.doi.org/10.1371/journal.pone.0179471] [PMID: 28614381]
- [11] Minavi B, Youssefi A, Quock R, et al. Evaluating the substantivity of silver diamine fluoride in a dentin model. *Clin Exp Dent Res* 2021; 7(4): 628-33. [http://dx.doi.org/10.1002/cre2.376] [PMID: 33291186]
- [12] A SR, Reddy N, Dinapadu S, Reddy M, Pasari S. Role of ozone therapy in minimal intervention dentistry and endodontics : A review. *J Int Oral Health* 2013; 5(3): 102-8. [PMID: 24155611]
- [13] Boch T, Tennert C, Vach K, Al-Ahmad A, Hellwig E, Polydorou O. Effect of gaseous ozone on enterococcus faecalis biofilm : An *in vitro* study. *Clin Oral Invest* 2016; 20(7): 1733-9. [http://dx.doi.org/10.1007/s00784-015-1667-1] [PMID: 26637463]
- [14] Case PD, Bird PS, Kahler WA, George R, Walsh LJ. Treatment of root canal biofilms of *Enterococcus faecalis* with ozone gas and passive ultrasound activation. *J Endod* 2012; 38(4): 523-6. [http://dx.doi.org/10.1016/j.joen.2011.12.020] [PMID: 22414842]
- [15] Gojanur S. Ozone therapy : An alternative to conventional therapy in pediatric dentistry. *Guident* 2015; 8.
- [16] Mei ML, Lo ECM, Chu CH. Arresting dentine caries with silver diamine fluoride: What's behind it? *J Dent Res* 2018; 97(7): 751-8. [http://dx.doi.org/10.1177/0022034518774783] [PMID: 29768975]
- [17] Briseño-Marroquín B, Ismael Y, Callaway A, Tennert C, Wolf TG. Antibacterial effect of silver diamine fluoride and potassium iodide against *E. faecalis*, *A. naeslundii* and *P. micra*. *BMC Oral Health* 2021; 21(1): 175. [http://dx.doi.org/10.1186/s12903-021-01531-1] [PMID: 33827520]
- [18] Koruyucu M, Bayram M, Kasımoğlu Y, Seymen F. Comparison of root canal length measurement methods in primary teeth. *Dentistry* 2018; 6: 8-16. [http://dx.doi.org/10.5195/d3000.2018.83]
- [19] Li M, Wong W, Xiong H, Chen K. *In vitro* antibacterial effects of photodynamic therapy against *Enterococcus faecalis* in root canals of deciduous teeth. *BMC Oral Health* 2022; 22(1): 554. [http://dx.doi.org/10.1186/s12903-022-02523-5] [PMID: 36457124]
- [20] Portenier I, Waltimo TMT, Haapasalo M. *Enterococcus faecalis*- the root canal survivor and 'star' in post-treatment disease. *Endod Topics* 2003; 6(1): 135-59. [http://dx.doi.org/10.1111/j.1601-1546.2003.00040.x]
- [21] Stojicic S, Shen Y, Haapasalo M. Effect of the source of biofilm bacteria, level of biofilm maturation, and type of disinfecting agent on the susceptibility of biofilm bacteria to antibacterial agents. *J Endod* 2013; 39(4): 473-7. [http://dx.doi.org/10.1016/j.joen.2012.11.024] [PMID: 23522539]
- [22] Kishen A, George S, Kumar R. *Enterococcus faecalis*-mediated biomaterialized biofilm formation on root canal dentine *in vitro*. *J Biomed Mater Res A* 2006; 77A(2): 406-15. [http://dx.doi.org/10.1002/jbm.a.30622] [PMID: 16444682]
- [23] Yu Q. Scavenging strategy for root canal infection. *Chin J Stomatol* 2018; 53: 381-5.
- [24] Baca P, Mendoza-Llamas ML, Arias-Moliz MT, González-Rodríguez MP, Ferrer-Luque CM. Residual effectiveness of final irrigation regimens on *Enterococcus faecalis*-infected root canals. *J Endod* 2011; 37(8): 1121-3. [http://dx.doi.org/10.1016/j.joen.2011.05.003] [PMID: 21763905]
- [25] Jaju S, Jaju PP. Newer root canal irrigants in horizon: A review *Int J dent* 2011; 2011. [http://dx.doi.org/10.1155/2011/851359]
- [26] Huth KC, Jakob FM, Saugel B, et al. Effect of ozone on oral cells compared with established antimicrobials. *Eur J Oral Sci* 2006; 114(5): 435-40. [http://dx.doi.org/10.1111/j.1600-0722.2006.00390.x] [PMID: 17026511]
- [27] Sivalingam VP, Panneerselvam E, Raja KV, Gopi G. Does topical ozone therapy improve patient comfort after surgical removal of impacted mandibular third molar? A randomized controlled trial. *J Oral Maxillofac Surg* 2017; 75(1): e1-51-9. [http://dx.doi.org/10.1016/j.joms.2016.09.014]
- [28] Naik SV, K R, Kohli S, Zohabhasan S, Bhatia S. Ozone: A biological therapy in dentistry : Reality or myth????? *Open Dent J* 2016; 10(1): 196-206. [http://dx.doi.org/10.2174/1874210601610010196] [PMID: 27386005]
- [29] Chu CH, Lo ECM, Lin HC. Effectiveness of silver diamine fluoride and sodium fluoride varnish in arresting dentin caries in Chinese pre-school children. *J Dent Res* 2002; 81(11): 767-70. [http://dx.doi.org/10.1177/0810767] [PMID: 12407092]
- [30] Yamaga R. Arrestment of caries of deciduous teeth with diamine silver fluoride. *Dent Outlook* 1969; 33: 1007-13.
- [31] Gomes-Filho JE, Silva FO, Watanabe S, et al. Tissue reaction to silver nanoparticles dispersion as an alternative irrigating solution. *J Endod* 2010; 36(10): 1698-702. [http://dx.doi.org/10.1016/j.joen.2010.07.007] [PMID: 20850681]
- [32] Hiraishi N, Yiu CKY, King NM, Tagami J, Tay FR. Antimicrobial efficacy of 3.8% silver diamine fluoride and its effect on root dentin. *J Endod* 2010; 36(6): 1026-9. [http://dx.doi.org/10.1016/j.joen.2010.02.029] [PMID: 20478459]
- [33] Selvig KA. Ultrastructural changes in human dentine exposed to a weak acid. *Arch Oral Biol* 1968; 13(7): 719-19. [http://dx.doi.org/10.1016/0003-9969(68)90090-3] [PMID: 4876376]
- [34] Suzuki T, Nishida M, Sobue S, Moriwaki Y. Effects of diamine silver fluoride on tooth enamel. *J Osaka Univ Dent Sch* 1974; 14: 61-72. [PMID: 4535096]
- [35] Al-Madi EM, Al-Jamie MA, Al-Owaid NM, Almohaimede AA, Al-

- Owid AM. Antibacterial efficacy of silver diamine fluoride as a root canal irrigant. *Clin Exp Dent Res* 2019; 5(5): 551-6. [<http://dx.doi.org/10.1002/cre2.222>] [PMID: 31687190]
- [36] Chaudhari DV, Shashikiran N, Maurya A, *et al.* Comparative evaluation of antimicrobial efficacy of sodium hypochlorite, silver diamine fluoride fluoride, chitosan and bioactive glass nanoparticles as root canal irrigants against the bacterial strain of enterococcus faecalis *In Vitro* Study. *J Clin Diagn Res* 2020; 14.
- [37] Huth KC, Quirling M, Maier S, *et al.* Effectiveness of ozone against endodontopathogenic microorganisms in a root canal biofilm model. *Int Endod J* 2009; 42(1): 3-13. [<http://dx.doi.org/10.1111/j.1365-2591.2008.01460.x>] [PMID: 19125975]
- [38] Silva EJNL, Prado MC, Soares DN, Hecksher F, Martins JNR, Fidalgo TKS. The effect of ozone therapy in root canal disinfection: A systematic review. *Int Endod J* 2020; 53(3): 317-32. [<http://dx.doi.org/10.1111/iej.13229>] [PMID: 31587303]
- [39] Narayanan LL, Vaishnavi C. Endodontic microbiology. *JCD* 2010; 13(4): 233-9. [PMID: 21217951]
- [40] Kaushik N, Rehani U, Agarwal A, Kaushik M, Adlakha V. Antimicrobial efficacy of endodontic irrigants against *Enterococcus faecalis* and *Escherichia coli*: An *in vitro* study. *Int J Clin Pediatr Dent* 2013; 6(3): 178-82. [<http://dx.doi.org/10.5005/jp-journals-10005-1214>] [PMID: 25206218]

